

BET bromodomain inhibitors abrogate cell cycle progression and induces apoptosis in Myc-induced mouse lymphoma cells without affecting MYC transcription



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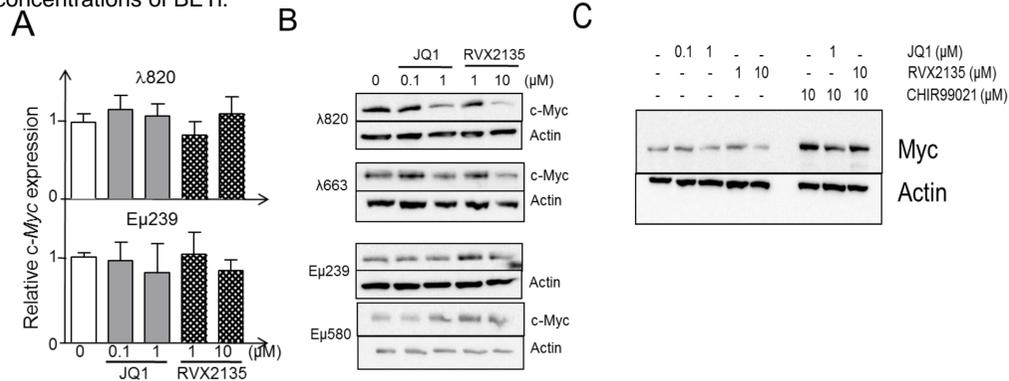
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Abstract

Overexpression of one of the three *MYC* genes is a hallmark of many human cancers. They encode transcription factors that regulate expression of a number of genes critical for tumor development. Conditional transgenic mouse models have shown that *Myc* inhibition causes tumor regression, *Myc* is therefore an attractive target for therapy and effective pharmacological *Myc* inhibition has been a long-standing goal in cancer research. Recent publications have shown that selective inhibitors of BET bromodomain and extra-terminal (BET) domain family of proteins, that bind to acetylated lysines on histones, show promise as potent anti-cancer drugs via down-regulation of the *MYC* oncogene. Here we confirm that two structurally different BET protein inhibitors (BETi), JQ1 (1) (proto-type) and RVX2135, inhibit the proliferation and induce apoptosis of lymphoma cells arising in *Myc*-transgenic mice. BETi have been reported to directly inhibit *MYC* transcription. Surprisingly, in our system BETi inhibition had no effect on *MYC* transcription, despite exhibiting broad transcriptional effects evident from expression profiling. Our data challenge the prevailing view that BETi operate primarily via suppression of *MYC* transcription in hematological malignancies (1-6). Instead we suggest that BET proteins have *Myc*-independent pleiotropic effect that should be exploited to treat a wide range of cancers. *Myc* still remains a challenging target for therapy to date.

Results

Figure 3. BETi does not suppress *Myc* transcription in *Myc* induced murine lymphoma cells. **A)** qRT-PCR analysis of *Myc* RNA expression in indicated cell lines treated with different concentrations of BETi. **B)** Western blot analysis of c-*Myc* protein levels. **C)** Western blot analysis of λ 820 cells treated for 24h with CHIR99021 (Selleck chemicals) in the presence or absence of the indicated concentrations of BETi.



Results

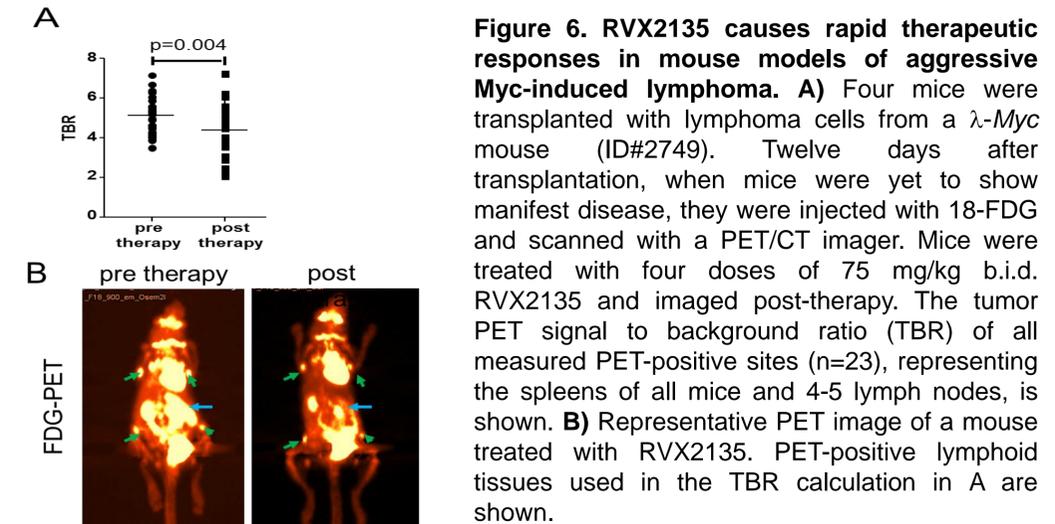


Figure 6. RVX2135 causes rapid therapeutic responses in mouse models of aggressive *Myc*-induced lymphoma. **A)** Four mice were transplanted with lymphoma cells from a λ -*Myc* mouse (ID#2749). Twelve days after transplantation, when mice were yet to show manifest disease, they were injected with 18-FDG and scanned with a PET/CT imager. Mice were treated with four doses of 75 mg/kg b.i.d. RVX2135 and imaged post-therapy. The tumor PET signal to background ratio (TBR) of all measured PET-positive sites (n=23), representing the spleens of all mice and 4-5 lymph nodes, is shown. **B)** Representative PET image of a mouse treated with RVX2135. PET-positive lymphoid tissues used in the TBR calculation in A are shown.

Results

Figure 1. Features of RVX2135, a novel BET inhibitor. **A)** Structure of the chemical scaffold from which RVX2135 was developed. **B)** FRET assay showing displacement of BET proteins by RVX2135.

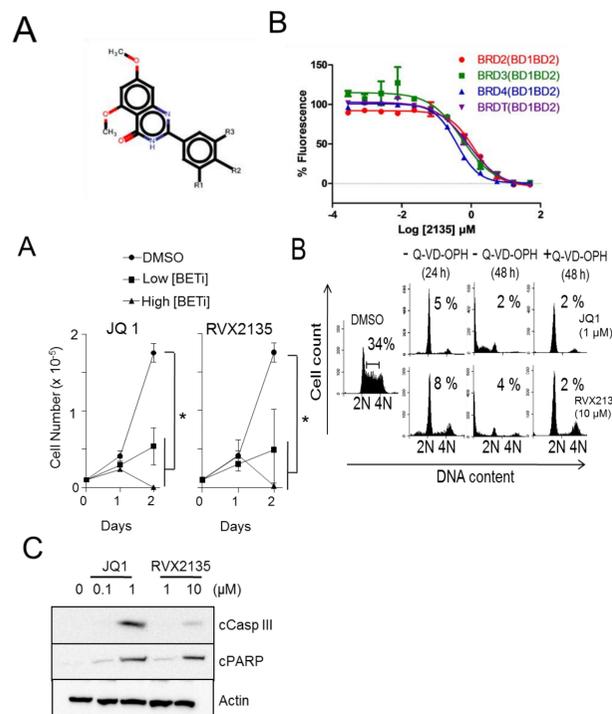


Figure 2. BET inhibitors induce cell cycle arrest and apoptosis of *Myc*-transgenic mouse lymphoma cells. **A)** λ 820 cells were treated for 48h with BETi and were counted. Low concentrations of BETi were defined as 100 nM JQ1 and 1 μ M RVX2135, whereas high concentrations as 1 μ M JQ1 and 10 μ M RVX2135. **B)** λ 820 cells were treated for 24h and 48h with BETi in the presence or absence of the pan-caspase inhibitor Q-VD-OPH. Cell cycle analysis by flow cytometry analysis of 7-AAD-stained cells, shown are cells in S-phase. **C)** Western blot analysis of λ 820 cells were treated for 24h with the indicated concentrations of BETi.

Figure 4. BETi induce broad transcriptional effects affecting several growth promoting gene signatures. **A)** Supervised hierarchical clustering of Illumina beadchip microarray data in λ 820 and Eµ239 cells treated with BETi. Shown are the 50 most down- and up-regulated genes (fold-change). **B)** Venn diagram of the genes down-regulated by treatment with λ 820 and Eµ239 cells with 1 μ M JQ1 for 24h. **C)** GSEA of genes co-regulated (down) by JQ1 in both λ 820 cells and Eµ239 cells. Shown are top transcription factors (TF) associated with the gene signatures.

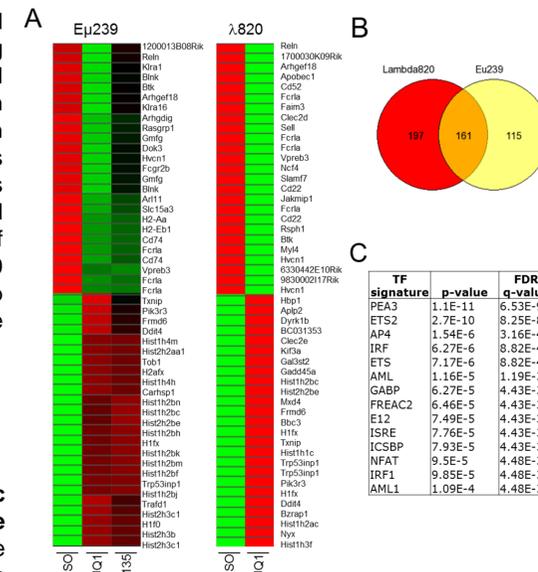
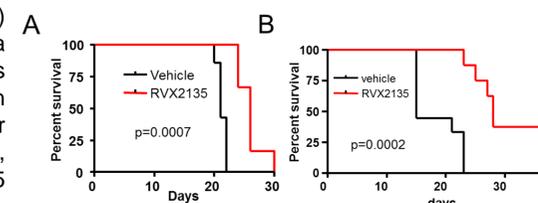


Figure 5. RVX2135 causes potent therapeutic responses in mouse models of aggressive *Myc*-induced lymphoma. **A)** λ 820 cells were transplanted into syngeneic B6 mice via tail vein injection. Four days after injection, mice were dosed with 75 mg/kg (b.i.d; five days per week) RVX2135 (n=6) or vehicle (n=7). **B)** A lymphoma arising in a λ -*Myc* mouse (ID#2749) was transplanted into recipient B6 mice via tail vein injection, accompanied by treatment with either vehicle or RVX-2135. Four days after injection, mice were dosed with 75 mg/kg b.i.d. RVX2135 (n=8) or vehicle (n=9).



Conclusions

RVX2135 blocks cell cycle progression and induces apoptosis in *Myc*-induced mouse lymphoma cells *in vitro* and in mice. Our data strongly suggests that the effect of BETi is likely not monogenic since several transcription factor networks are altered. Previous studies in hematological malignancies and some solid tumors have suggested that the main target of inhibition by BETi is *MYC* (1-6). By using GEM models of lymphoma it has here been possible to uncouple BETi effects on *Myc* transcription from the effects of BETi on other transcription factor networks. Interestingly, despite maintaining c-*Myc* expression, the murine B-cell lymphoma cells are as sensitive to BETi as the human lymphoma cells previously studied (2-6). Therefore the fact that there is no genetic rescue in the mouse model demonstrates that BETi block lymphoma cell proliferation and induces apoptosis independent of inhibiting *Myc* transcription in B-cell lymphoma. This finding underscores the complexity of epigenetic regulation of transcription factor networks in cancer cells.

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References

- Filippakopoulos P, Qi J, Picaud S, Shen Y, Smith WB, Fedorov O, et al. Selective inhibition of BET bromodomains. *Nature*. 2010;468:1067-73.
- Delmore JE, Issa GC, Lemieux ME, Rahl PB, Shi J, Jacobs HM, et al. BET bromodomain inhibition as a therapeutic strategy to target c-Myc. *Cell*. 2011;146:904-17.
- Zuber J, Shi J, Wang E, Rappaport AR, Herrmann H, Sison EA, et al. RNAi screen identifies Brd4 as a therapeutic target in acute myeloid leukaemia. *Nature*. 2011;478:524-8.
- Zhao X, Lwin T, Zhang X, Huang A, Wang J, Marquez VE, et al. Disruption of the MYC-miRNA-EZH2 loop to suppress aggressive B-cell lymphoma survival and clonogenicity. *Leukemia*. 2013.
- Tolani B, Gopalakrishnan R, Punj V, Matta H, Chaudhary PM. Targeting Myc in KSHV-associated primary effusion lymphoma with BET bromodomain inhibitors. *Oncogene*. 2013.
- Ott CJ, Kopp N, Bird L, Paranal RM, Qi J, Bowman T, et al. BET bromodomain inhibition targets both c-Myc and IL7R in high-risk acute lymphoblastic leukemia. *Blood*. 2012;120:2843-52.